

Impact of the Prone Position in an Animal Model of Unilateral Bacterial Pneumonia Undergoing Mechanical Ventilation

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ABSTRACT

Background: The prone position (PP) has proven beneficial in patients with severe lung injury subjected to mechanical ventilation (MV), especially in those with lobar involvement. We assessed the impact of PP on unilateral pneumonia in rabbits subjected to MV.

Methods: After endobronchial challenge with *Enterobacter aerogenes*, adult rabbits were subjected to either “adverse” (peak inspiratory pressure = 30 cm H₂O, zero end-expiratory pressure; n = 10) or “protective” (tidal volume = 8 ml/kg, 5 cm H₂O positive end-expiratory pressure; n = 10) MV and then randomly kept supine or turned to the PP. Pneumonia was assessed 8 h later. Data are presented as median (interquartile range).

Results: Compared with the supine position, PP was associated with significantly lower bacterial concentrations within the infected lung, even if a “protective” MV was applied (5.93

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Received from UMR 1347, pôle Microbiologie Environnementale et Risque Sanitaire, Université de Bourgogne, Faculté de Médecine, Dijon, France, and the Department of Microbiology and Molecular Medicine, Faculty of Medicine, University of Geneva, Geneva, Switzerland. Submitted for publication February 11, 2012. Accepted for publication January 11, 2013. Supported by the “Bourse Qualité Recherche” from the Université de Bourgogne, Dijon, France, obtained in 2007 in addition to institutional sources. The first two authors contributed equally to this work.

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What We Already Know about This Topic

- Prone positioning can improve gas exchange in acute lung injury patients

What This Study Tells Us That Is New

- In anesthetized animals with unilateral bacterial-induced lung injury subjected to nonprotective mechanical ventilation, prone positioning (compared to supine positioning) decreases the bacterial concentration in the infected lungs and attenuates lung and systemic inflammation

[0.34] vs. 6.66 [0.86] log₁₀ cfu/g, respectively; $P = 0.008$). Bacterial concentrations in the spleen were also decreased by the PP if the “adverse” MV was used (3.62 [1.74] vs. 6.55 [3.67] log₁₀ cfu/g, respectively; $P = 0.038$). In addition, the noninfected lung was less severely injured in the PP group. Finally, lung and systemic inflammation as assessed through interleukin-8 and tumor necrosis factor- α measurement was attenuated by the PP.

Conclusions: The PP could be protective if the host is subjected to MV and unilateral bacterial pneumonia. It improves lung injury even if it is utilized after lung injury has occurred and nonprotective ventilation has been administered.

MECCHANICAL ventilation (MV) is widely used in critically ill patients with respiratory failure. Cumulative evidence suggests, however, that MV could be harmful for the lung. Thus, both overdistension of the airways and intratidal alveolar cyclic opening and closing are likely to cause tissue damage, called ventilator-induced lung injury (VILI).¹ In addition, lung stretch could lead to the release of inflammatory cytokines, thereby causing additional injury, particularly through the recruitment of polymorphonuclear neutrophils mediated by interleukin (IL)-8.²⁻⁴ Finally, some experimental findings have increased the possibility that the ability of the host to keep bacterial growth in check could be hampered by the mechanical forces applied to the lung

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subjected to MV.^{5–9} Although these VILI features were primarily described in animals with preinjured lungs subjected to ventilator settings never applied to the patients (*i.e.*, very large tidal volumes [V_T]), more recent data support the hypothesis that mild but consistent tissue damages as well as activation of pulmonary inflammation could appear with usual MV parameters.^{10,11}

Nonetheless, limiting alveolar strain by reducing V_T and preventing alveolar expiratory collapse through the application of positive end-expiratory pressure (PEEP) have proven to be lung protective in animals as well as in patients with severe lung injury.^{8,12,13} However, it has been shown that despite “protective” MV, both injury and inflammation were unevenly distributed within the lung.^{14,15} Thus, although the nondependent areas tend to be over distended, the dependent lung regions are likely to be poorly aerated. Lobar injury caused by bacterial pneumonia could worsen these features.¹⁶ In addition, although some experimental studies have shown that “protective” MV strategy was likely to improve the outcome of pneumonia, if compared with an “adverse” one (high V_T plus zero end-expiratory pressure), some issues remain unsolved.^{17–21} First, despite being considered “protective” MV is not necessarily safe as it could worsen tissue damage and increase bacterial growth in animals with pneumonia.^{5,6} In addition, setting the right level of PEEP remains a matter of great concern. Actually, raising the PEEP could lead to lung overdistension in patients with lobar acute respiratory distress syndrome.^{22,23} Animal models have shown that high PEEP was associated with overdistension around pneumonia foci and within the contralateral lung as well, thereby promoting lung inflammation and pulmonary-to-systemic bacterial translocation.^{24,25}

The prone position (PP) has been shown to improve gas exchange and lung mechanics in patients with acute respiratory distress syndrome.²⁶ This seems especially true in patients with lobar acute respiratory distress syndrome.²⁷ However, available animal models of diffuse lung injury showed that the PP could safely improve alveolar recruitment through a better tidal ventilation distribution within the lung, thereby delaying if not attenuating VILI features.^{28–31} However, it has not been tested in models of lung injury with lobar involvement so far. We hypothesized, therefore, that the PP could reduce the severity of unilateral bacterial pneumonia in rabbits subjected to either “adverse” or “protective” MV.

Materials and Methods

Objectives

An experimental study was carried out in order to assess the impact of PP on bacterial pneumonia with two main endpoints:

- the severity of the unilateral pneumonia;
- the VILI features within the noninfected contralateral lung.

Animals

Male New Zealand white rabbits (body weight, 2.7–3.0 kg) were obtained from Elevage scientifique des Dombes (Romans, France). These animals were not immunosuppressed and were free of both virus antibodies and specific pathogens. They were placed in individual cages and had free access to water and were fed in accordance with current recommendations mentioned in the *Guide for the Care and Use of Laboratory Animals*, National Institutes of Health No. 92–23, revised 1985. The Dijon Faculty of Medicine Ethical Committee approved the experimental protocol. A central venous catheter was surgically inserted into every rabbit the day before MV.

Experimental Protocol

The animals were intubated as previously described.⁵ Briefly, during general anaesthesia provided by iterative intravenous injections of propofol, a cuff tube of 3.0 mm was orally inserted into the trachea under view control. The animal was put in the supine position (SP) and connected to a pressure-controlled respirator. During the first 30 min, the V_T was set at 8 ml/kg with zero end-expiratory pressure, a respiratory rate of 30 breaths/min and an inspired fraction of oxygen (FI_{O_2}) of 0.5 (fig. 1). Throughout the experiment, a continuous infusion of ketamine ($1 \text{ mg}\cdot\text{kg}^{-1} \text{ h}^{-1}$) and pancuronium bromide ($0.3 \text{ mg}\cdot\text{kg}^{-1} \text{ h}^{-1}$), was given. Hydration with isotonic serum was provided intravenously so that the rabbits remained at a constant weight.

Enterobacter aerogenes pneumonia was induced as previously described.^{5,24} Briefly, one $10 \log_{10}$ cfu inoculum of a clinical strain of *E. aerogenes* was instilled through the left stem bronchus. According to previous data, it takes 3 h for histologically proven lobar pneumonia to develop.

In the first set of experiments, the impact of PP was tested in animals subjected to “adverse” MV strategy. Thus, 1 h after bacterial challenge, the peak inspiratory pressure was set at 30 cm H_2O and the animals were randomly kept in the SP ($n = 4$) or turned to the PP ($n = 5$). The animals were subjected to MV for 8 h before being killed by an overdose of thiopental.

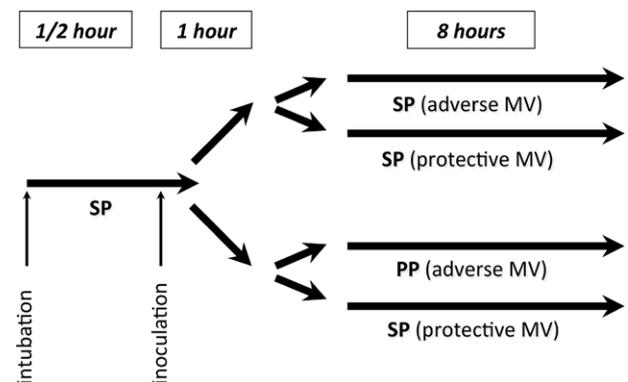


Fig. 1. Timeline representation of the experimental protocol. MV = mechanical ventilation; PP = prone position; SP = supine position.

In the second set of experiments, to match with the clinical practice, the animals were subjected to “protective” MV ($V_T = 8$ ml/kg; PEEP = 5 cm H₂O), once inoculated and kept in either the SP ($n = 5$) or turned to the PP ($n = 5$), similarly to the first set of experiments.

Assessment of Respiratory System Compliance and Other Physiological Measurements

Inspiratory pressure–volume curves were constructed *post-mortem* according to the supersyringe method. Respiratory system compliance (C_{RS}) was deduced from the slope of the linear portion of the pressure–volume curve.

An arterial catheter was inserted in most of the animals subjected to “adverse” MV for blood sampling and blood pressure monitoring to ascertain the safety of our protocol ($n = 7$). Thus, arterial blood gases and lactate levels were measured at randomization and before sacrifice. Variations in P_{aO_2} , P_{aCO_2} from H0 (randomization to the SP or the PP group) to H8 (sacrifice) were thus measured as surrogates for the alveolar recruitment. Arterial blood lactate was measured to ascertain the safety of our “adverse” MV.

It is worth noting that none of the animals subjected to “protective” MV was thus monitored. Actually, we had previously checked the safety of these settings as well as the lack of hypoxemia and hyperlactatemia in mechanically ventilated rabbits with this type of pneumonia.^{5,24}

Evaluation of Unilateral Pneumonia

The animals were exsanguinated by venous puncture. Autopsies were carried out and the lungs and spleen were harvested aseptically.

Each lung was isolated and homogenized in sterile water. Tenfold dilution cultures were then performed. The mean bacterial concentration of the infected lung was calculated according to the lung weight. The spleen of each rabbit was also crushed and cultured since a positive *E. aerogenes* spleen culture was considered a marker of bacteremia.

The remaining lung and spleen homogenates were then frozen, batched, and stored at -80°C until tissue concentrations of cytokines were measured. Accordingly, IL-8 and tumor necrosis factor- α were assessed using a rabbit-specific enzyme-linked immunosorbent assay following the manufacturer’s instructions (Euromedex®, Strasbourg, France).

Blood samples were obtained before bacterial inoculation (H0), at H1 and H8 only in animals in the second set of experiments (*i.e.*, “protective” MV), to assess systemic inflammation according to the position. Thus, IL-8 and tumor necrosis factor- α blood concentrations were measured using the above-mentioned enzyme-linked immunosorbent assay test.

Assessment of VILI and Inflammation within the Noninfected Lung

For microscopic examination, approximately 1 cm³ of tissue was fixed in formalin and embedded in paraffin. Four-micrometer sections were obtained and stained with hematoxylin-eosin. A pulmonary pathologist blinded to the treatment group examined 10 fields of each section and an injury score was calculated as previously described.²⁸ Briefly, lung injury assessment was based on the degree of neutrophilic infiltration, hemorrhage, and oedema. Lung injury was considered absent (0), mild (1), moderate (2), or severe (3). In addition, the presence or absence of hyaline membranes as well as emphysema-like lesions was systematically sought.

Another lung sample was harvested for RNA extraction using the GenElute kit (Sigma®, Dorset, United Kingdom). RNA extraction was performed using the RNA GenElute kit accordingly. Complementary DNA was obtained by reverse transcription using random primers, RNAs in treatment, and ImProm II reverse transcriptase (Promega®, Madison, WI). Quantitative polymerase chain reaction was performed using the IQ5 thermocycler (Biorad®, Hercules, CA) and the IQ Sybergreen Supermix (Biorad®) and rabbit-specific primers, designed using Primer3 (version 0.4.0),** and the rabbit (*Oryctolagus cuniculus*) sequence database.†† Melting curves were plotted to check the specificity of the amplifications. The following primers were used: *rGapdh* forward: 5′-ATG TTT GTG ATG GGC GTG AAC C-3′, reverse: 5′-CCC AGC ATC GAA GGT AGA GGA-3′; *rIl-8* forward: 5′-AAC CTT CCT GCT GCT TCT GA-3′, reverse: 5′-TCT GCA CCC ACT TTT TCC TTG-3′. The results were expressed as expression levels normalized to a reference gene (*rGapdh*). The corresponding protein assessment could not however be achieved since all of the remaining tissue had to be used for gravimetric evaluation of the lung as described below.

The remaining fresh tissue was weighed to measure the lung wet weight (WW) and warmed to 37°C until desiccation before recording the dry weight (DW). The WW to DW ratio (WW/DW) was then calculated as a surrogate for lung permeability oedema. Previously, healthy rabbits with normal lung tissue ($n = 4$) were killed and used as controls.

Statistical Analysis

Data are presented as median (interquartile range) except otherwise stated. The Mann–Whitney U test was used to compare continuous variables between groups. All tests were two-tailed. A $P \leq 0.05$ was considered significant. The Statview software was used (SAS Institute, Cary, NC) for all analysis.

Results

C_{RS} Gas Exchange and Hemodynamics

C_{RS} was measured *postmortem* in all rabbits with pneumonia (table 1). C_{RS} was greater in animals that had undergone “protective” MV than in the others but remained unchanged by the body position. In addition, C_{RS} was improved by PP

** Available at: www.primer3plus.com. Accessed March 5, 2008.

†† Available at: www.ensembl.org. Accessed March 5, 2008.

Table 1. Physiological Parameters Recorded in Rabbits with Unilateral *Enterobacter aerogenes* Pneumonia Submitted to Adverse MV in either SP or PP

	C_{RS} , ml/cm H ₂ O	$\Delta P_{aO_2}/F_{iO_2}$, mmHg	ΔP_{aCO_2} , mmHg	MAP, mmHg	Heart Rate, beat/min	Δ (Lactate), mmol/l
Adverse MV						
SP (n = 3)	2.8 (1.4)	-69 (77)	-11 (4)	79.0 (12.7)	276 (30)	2.1 (3.1)
PP (n = 4)	5.2 (1.1)*	235 (244)*	-11 (2)	75.3 (4.7)	262 (24)	-0.4 (0.7)*
Protective MV						
SP (n = 5)	13.5 (0.9)					
PP (n = 5)	11.4 (3.8)					

Results are presented as median \pm IQR. The delta values mean variation from H0 (randomization to the SP or the PP group) to H8 (sacrifice).

* $P < 0.05$ between SP and PP.

C_{RS} = respiratory system compliance; F_{iO_2} = inspired fraction of oxygen; IQR = interquartile range; MAP = mean arterial pressure; MV = mechanical ventilation; PP = prone position; SP = supine position.

if “adverse” MV strategy was applied (2.8 [1.4] *vs.* 5.2 [1.1] ml/cm H₂O; $P = 0.001$).

Differences were also observed regarding gas exchange in the animals subjected to “adverse” MV in which blood gases could be obtained. An increase of 235 (244) mmHg in the P_{aO_2}/F_{iO_2} ratio was recorded in the PP group, whereas a decrease of 69 (77) mmHg was measured in the supine animals ($P = 0.04$). No difference was seen regarding blood arterial pressure and heart rate measured just before sacrifice. However, the blood lactate concentration in the SP increased throughout the experiment while it remained within normal range in the PP (Δ [lactate] = 2.1 [3.1] *vs.* -0.4 [0.7] mmol/l, respectively; $P = 0.034$) (table 1).

Assessment of Unilateral Pneumonia

All of the animals but one (SP) survived until they were killed 8 h after having being turned prone or kept supine. The size of the *E. aerogenes* inoculum in the SP and PP groups was found to be comparable, regardless of the MV strategy (data not shown).

Quantitative lung culture results showed high mean concentrations of *E. aerogenes* 8 h after inoculation (fig. 2). Lower concentrations were found in the PP group than in the SP group (8.38 [0.91] *vs.* 9.81 [0.52] \log_{10} cfu/g of tissue, respectively; $P = 0.002$) when an “adverse” MV strategy was used. Although “protective” MV decreased the bacterial pulmonary burden regardless of the position, the lowest bacterial concentrations within this group were found in animals in the PP (5.93 [0.34] *vs.* 6.66 [0.86] \log_{10} cfu/g, respectively; $P = 0.008$). When the lung inflammatory response was considered, the PP was associated with the release of smaller amounts of IL-8, regardless of the MV strategy (fig. 3). In contrast, although a “protective” MV significantly decreased pulmonary concentrations of tumor necrosis factor- α regardless of the position (2155 [830] *vs.* 3955 [1412] pg/g of tissue, respectively; $P = 0.014$), the PP did not decrease cytokine release further.

The extrapulmonary impact of pneumonia was also assessed. Like the lung, spleen culture results showed

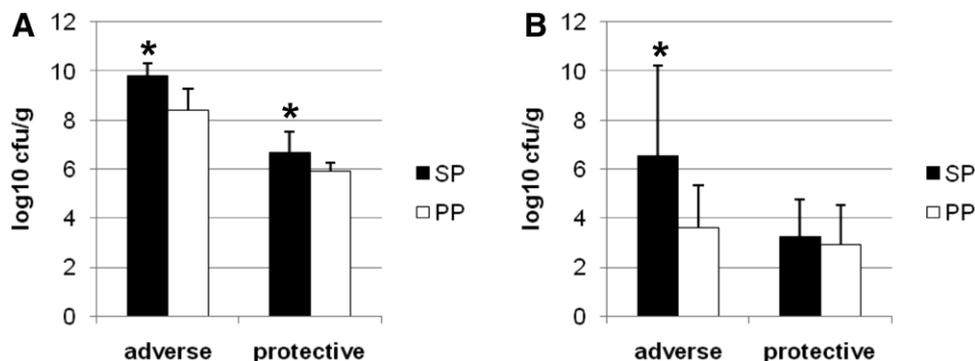


Fig. 2. Bacterial burden (infected lung, [A]; spleen, [B]) in animals with *Enterobacter aerogenes* pneumonia submitted to an adverse (n = 9) or a protective (n = 10) mechanical ventilation (MV) in supine position (SP), or turned from supine to prone position (PP). There is a bacterial overgrowth in the lung of the animals from the SP group, whenever the MV is protective or not. The pulmonary-to-systemic translocation is increased by SP in the only animals submitted to an adverse MV strategy, as reflected by the larger amount of bacteria recovered from the spleen culture in this group. Results are expressed as median \pm IQR. * $P < 0.05$ between SP and PP groups. IQR = interquartile range.

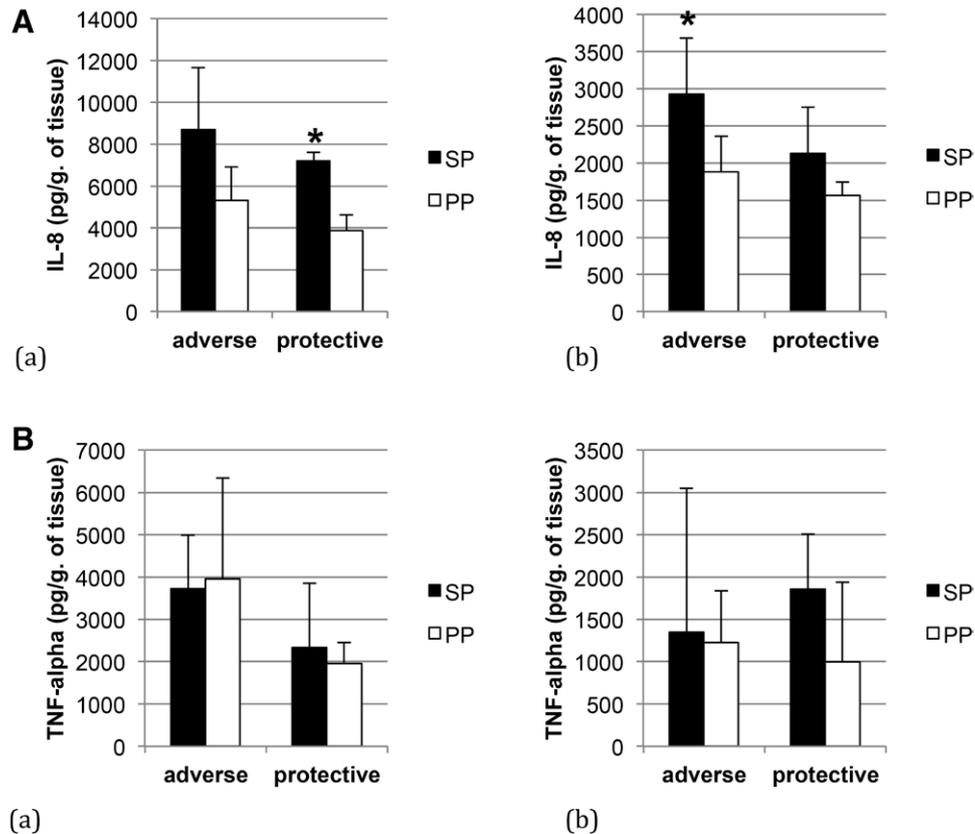


Fig. 3. Interleukin (IL-8) (A) and tumor necrosis factor- α (TNF- α) (B) concentrations in the infected lung (a), and the spleen (b) of rabbits with *Enterobacter aerogenes* unilateral pneumonia according to the mechanical ventilation strategy (i.e., “adverse” or “protective”), and the position (i.e., supine position [SP] or prone position [PP]). Results are expressed as median \pm IQR. It is worth noting that IL-8 release was decreased within the pulmonary and the systemic compartment in the animals turned into PP. In contrast, no significant difference was seen regarding TNF- α . * $P < 0.05$ between SP and PP groups. IQR = interquartile range.

lower bacterial concentrations in the animals turned to the PP than in those kept supine within the “adverse” MV group (3.62 [1.74] vs. 6.55 [3.67] \log_{10} cfu/g, respectively; $P = 0.038$). However, pulmonary-to-systemic translocation of *E. aerogenes* was not lower in prone than in supine animals subjected to “protective” MV (2.93 [1.59] vs. 3.24 [1.51] \log_{10} cfu/g, respectively; $P = 0.684$). The systemic inflammatory response was evaluated by the measurement of two of its key mediators within the blood compartment (fig. 3). Although concentrations of tumor necrosis factor- α in the spleen were similar in both groups regardless of the MV strategy, lower amounts of IL-8 were found in the PP than in the SP group when the rabbits underwent “adverse” MV (1910 [784] vs. 3005 [1290] pg/g of tissue, respectively; $P = 0.038$). In the animals subjected to “protective” MV, a statistically nonsignificant lower IL-8 release was found in the PP group (1561 [274] vs. 1930 [1022] pg/g of tissue, respectively; $P = 0.124$). Similarly, blood concentrations of IL-8 rose more slowly with time when the rabbits were turned to the PP than when they were kept supine (fig. 4).

Assessment of Lung Injury and Inflammation in the Noninfected Lung

Microscopic examination revealed that lung injury within the noninfected lung depended on both the MV strategy and body position (fig. 5). As expected, “protective” MV was associated with less tissue damage. The histologic scores tended to be greater in animals from the SP group than in those from the PP group, regardless of the MV strategy (table 2). Thus, there was an obvious loss of aeration within the lower lobe in the animals kept supine while the airspaces within the upper lobe appeared enlarged, especially if the MV was “adverse.” In contrast, lung aeration in the upper and lower lobes of the animals ventilated in the PP was quite similar. In addition, features like hyaline membranes and emphysema-like lesions were mainly seen in the animals kept supine.

However, no difference was found according to the body position regarding the WW/DW ratio when the animals were ventilated adversely. This suggests the formation of comparable amounts of permeability oedema in the two groups since far lower values were measured in the healthy rabbits (table 2). In contrast, the WW/DW ratio was significantly lower in the PP than in the SP when “protective” MV was applied.

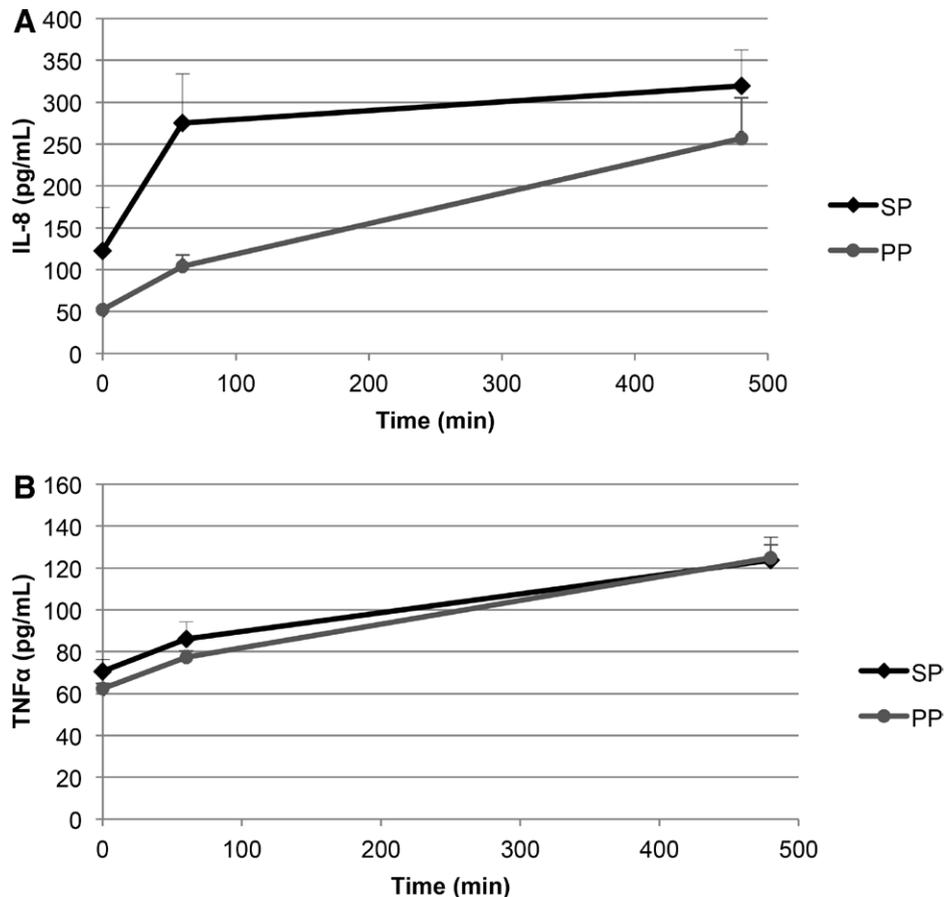


Fig. 4. Interleukin (IL)-8 (A) and tumor necrosis factor- α (TNF- α) (B) serum concentrations in rabbits with *Enterobacter aerogenes* unilateral pneumonia submitted to the “protective” mechanical ventilation according to the position (*i.e.*, supine position [SP] or prone position [PP]). Results are presented as median \pm IQR. A statistically nonsignificant increased of the IL-8 levels within the blood compartment in the animals in PP compared to those kept supine. IQR = interquartile range.

The evaluation of pulmonary inflammation within the noninfected lung was based on IL-8 gene expression. We observed that there was a stronger induction of IL-8 gene expression in the noninfected lung from SP animals than in those from PP animals regardless of the MV strategy (fig. 6).

Discussion

In the current study, we showed that turning animals from the supine to the PP was likely to improve the features of *E. aerogenes* unilateral pneumonia regardless of the MV strategy (*i.e.*, “adverse” or “protective”). Most of all, bacterial concentrations within both the instilled lung and the spleen were found to be lower in rabbits from the PP group than in those kept supine. In addition, IL-8 concentrations, a powerful chemoattractant for polymorphonuclears, were greater in the infected lung of the supine animals, as well as in an extrapulmonary organ, the spleen, suggesting that both lung and systemic inflammation were blunted by the PP. Taken together, these findings suggest that in our model, the PP improved lung bacterial clearance, reduced pulmonary-to-systemic translocation of bacteria, and mitigated the host inflammatory response. In addition, we showed that in this model of

lobar lung injury, the PP was likely to diminish the damage inflicted by MV to the noninfected lung and to modulate inflammation as well. Moreover, our findings illustrate the hypothesis that although pneumonia is less severe when using clinically relevant ventilator settings, further improvements could be obtained by turning the animals to the PP.

The best way to ventilate patients with lung injury remains a matter of concern. Improving gas exchange and optimizing alveolar recruitment are key issues in this setting. However, animal models of ventilator-associated pneumonia similar to ours have shown that pneumonia could induce airway overdistension within the infected as well as the “healthy” lung in animals subjected to MV.^{16,32} This deleterious effect of MV has been attributed to the loss of aeration within the infected pulmonary area, which could not be easily recruited by positive pressure. The magnitude of the local inflammatory response could account for this decrease in lung compliance.³³ As a result, although protective if set at a moderate level, PEEP may become harmful if higher levels are reached since it could create lung overdistension.^{25,34}

Prone positioning has been proposed primarily as an efficient way to improve gas exchange in patients with the

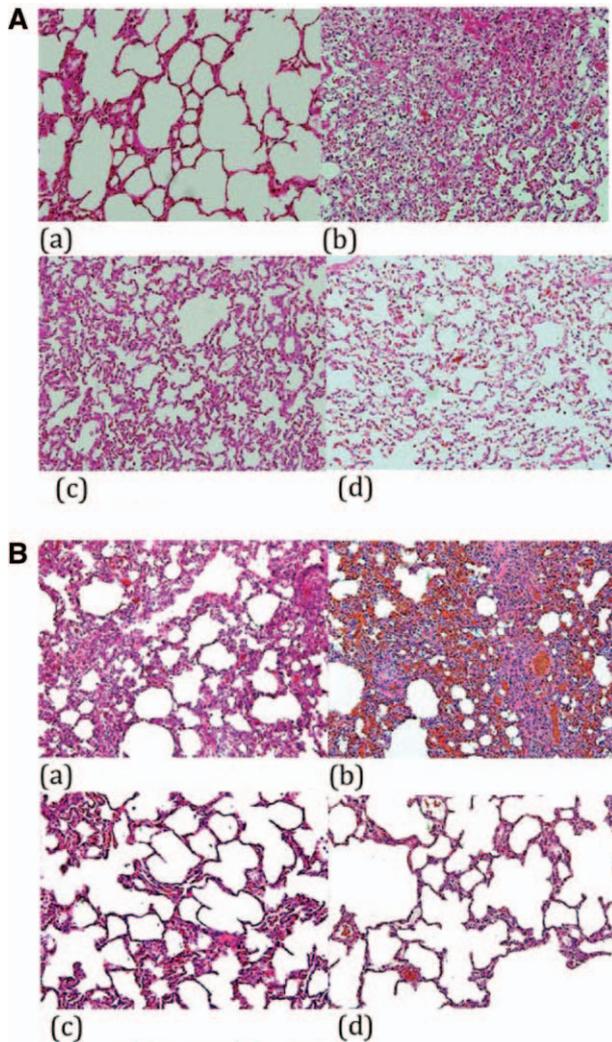


Fig. 5. Light photomicrographs of the noninfected lungs from animals with *Enterobacter aerogenes* pneumonia submitted to either an adverse (A) or a protective (B) mechanical ventilation (MV) in supine position (*upper lobe, a; lower lobe, b*), or prone position (PP) (*upper lobe, c; lower lobe, d*). (Original magnification, $\times 10$; hematoxylin-eosin). Neutrophil infiltration was greater in the animals submitted to an adverse MV, especially if kept supine. In addition, SP lead to a significant loss of aeration within the lower lobe while the tidal volume looked evenly distributed in the animals turned into PP, regardless of the MV strategy.

most severe forms of acute lung injury.³⁵ More recently, it has been shown that the PP allows better distribution of lung inflation along the craniocaudal axis through improvement in respiratory system compliance, together with lung recruitment.^{27,36} Interestingly, some authors have shown that those patients in whom compliance of the respiratory system improved once turned to the PP were more likely to present with lobar lung injury including pneumonia.²⁷ In addition, some recently published clinical studies showed that PP could either prevent ventilator-associated pneumonia or improve the outcome of existing ventilator-associated

pneumonia, whenever patients had lung injury before pneumonia.^{37–39} Altogether, these findings suggest that PP could be of particular interest in the setting of lobar lung injury including bacterial pneumonia. Our results provide additional insights that are likely to improve our understanding of such data since they draw a link between the well-known beneficial effects of the PP on lung mechanics, and a possible improvement in the host response against bacterial infection. We were, however, unable to determine the underlying mechanisms as far as the host innate immunity is concerned. Further studies are necessary since as shown by our group and others the toll-like receptors pathway could be altered by the unusual mechanical stretch applied to the lung subjected to MV.^{4,40,41} One could only speculate that the PP attenuates lung distension leading in turn to the release of smaller amount of IL-8, thereby protecting the host from the detrimental effects of an overwhelming inflammatory response.

In addition to such beneficial effects of the PP on the infected lung regarding bacterial clearance and the inflammatory response, we showed that the PP was likely to reduce lung injury within the noninfected lung. The decrease in PCO_2 subsequent to the increase in tidal ventilation (*i.e.*, when peak inspiratory pressure was set from 12 to 30 cm H_2O) was the same regardless of the body position, indicates that the PP did not necessarily increase the end-expiratory volume, as reported previously in acute respiratory pressure patients.^{35,42} However, strikingly, the distribution of lung aeration appeared to be different in the SP and the PP groups. In the SP group, there was a marked loss of aeration within the lower lobe whereas the airspaces of the upper lobe were enlarged. This resulted in the presence of emphysema-like lesions. In addition, the presence of hyaline membranes, one of the hallmarks of VILI was encountered exclusively in the animals that were ventilated supine. Similar findings were obtained when the “protective” ventilator settings were applied, although tissue injury was less severe. Interestingly, IL-8 gene expression was markedly higher in the noninfected lung of the supine animals than in those turned to the PP, and this was independent of the MV strategy. This could be considered a surrogate for biotrauma induced by MV, thereby indicating a greater level of lung distension in the SP group.⁴³ Surprisingly, there was no difference between SP and PP animals subjected to the “adverse” MV regarding the formation of lung permeability oedema within the noninfected lung as assessed by gravimetric measurements. We can only hypothesize that its distribution within the lung was different between groups since our approach (*i.e.*, WW/DW measurement) provided only an overall assessment. However, as expected, V_T reduction together with PEEP probably decreased the WW/DW ratio. In addition, we showed that further improvement was achieved by turning the animals in the PP. Altogether, these findings suggest that the PP improves V_T distribution

Table 2. Lung Injury Assessment within the Noninfected Lung of Rabbits with Unilateral *Enterobacter aerogenes* Pneumonia Submitted to Adverse or Protective MV in Either SP or PP

	Histologic Score	Hyaline Membranes	Emphysema-like Lesions	WW/DW
Control (n = 4)	—	—	—	1.7 (0.4)*
Adverse MV				
SP (n = 4)	4.0 (3.5)	3/4	2/4	5.3 (1.4)
PP (n = 5)	1.5 (2.7)	0/5	1/5	5.2 (1.4)
Protective MV				
SP (n = 5)	2.2 (1.0)	4/5	3/5	3.8 (1.3)
PP (n = 5)	1.1 (0.45)	1/5	0/5	2.7 (0.8)†

Results are presented as median \pm IQR. Healthy spontaneously breathing animals were used as controls.

* $P < 0.05$ between C and either PP or SP. † $P < 0.05$ between SP and PP.

DW = dry weight; IQR = interquartile range; MV = mechanical ventilation; PP = prone position; SP = supine position; WW = wet weight.

within the lung and in turn reduces airspace overdistension thus preventing VILI, as already demonstrated in animal models of diffuse lung injury.^{28–30}

Our study has several limitations. First, any extrapolation of our findings should be done very cautiously since small animals are known to be more prone to VILI than larger ones. Moreover, the MV settings used in the “adverse MV” group is far from the clinical practice. The way pneumonia was induced as well as the short duration of MV were also specific to our model and further experimental studies are necessary before our findings can be generalized. Second, one could argue that bacterial growth was better

controlled in the PP because of improved drainage of bronchial secretions regardless of any alteration of lung strain.⁴⁴ However, this could hardly account for the worsening of the lung injury within the contralateral lung. Third, the hemodynamic assessment was not performed extensively. As a result, we cannot exclude the possibility that the lower pulmonary-to-systemic translocation of both bacteria and mediators was subsequent to a drop in cardiac output in the PP group. However, previous clinical and experimental studies failed to show any difference regarding this point.^{26,30,36,45} Finally, we should acknowledge the small size of our experimental groups and the lack of any *a priori* calculation. Our study is therefore underpowered, making statistics difficult to interpret.

Conclusion

In a model of lobar lung injury, the PP may not only improve lung mechanics and blood oxygenation, but also enhance antibacterial defences and mitigate inflammation while protecting the contralateral lung.

The authors are grateful to Sonia Da Silva and Davy Hayez (Lab Technicians, Vivexia® Biotech, Dijon, France) for their technical contribution, and Amandine Bataille (Lab Technician, Cellimap, Université de Bourgogne, Dijon, France) for the histological section preparation.

References

- Dreyfuss D, Saumon G: Ventilator-induced lung injury: Lessons from experimental studies. *Am J Respir Crit Care Med* 1998; 157:294–323
- Kawano T, Mori S, Cybulsky M, Burger R, Ballin A, Cutz E, Bryan AC: Effect of granulocyte depletion in a ventilated surfactant-depleted lung. *J Appl Physiol* 1987; 62:27–33
- Pugin J, Dunn I, Jolliet P, Tassaux D, Magnanat JL, Nicod LP, Chevrolet JC: Activation of human macrophages by mechanical ventilation *in vitro*. *Am J Physiol* 1998; 275:L1040–50
- Charles PE, Tissières P, Barbar SD, Croisier D, Dufour J, Dunn-Siegrist I, Chavanet P, Pugin J: Mild-stretch mechanical ventilation upregulates toll-like receptor 2 and sensitizes the lung to bacterial lipopeptide. *Crit Care* 2011; 15:R181

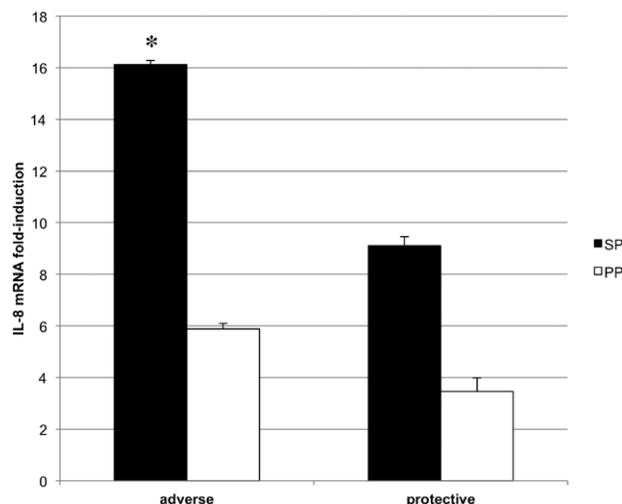


Fig. 6. Interleukin (IL)-8 messenger RNA levels in the noninfected lung of rabbits with *Enterobacter aerogenes* unilateral pneumonia according to the mechanical ventilation (MV) strategy (*i.e.*, “adverse” or “protective”), and the position (*i.e.*, supine position [SP] or prone position [PP]). All values are reported as fold increase compared with control spontaneously breathing healthy animals normalized to one. Results are expressed as mean \pm SD. There was a decrease of the IL-8 gene expression within the noninfected lung of the animals turned into PP, regardless of the MV settings. * $P < 0.05$ between SP and PP groups.

5. Charles PE, Piroth L, Desbiolles N, Lequeu C, Martin L, Portier H, Chavanet P: New model of ventilator-associated pneumonia in immunocompetent rabbits. *Crit Care Med* 2002; 30:2278–83
6. Marquette CH, Wermert D, Wallet F, Copin MC, Tonnel AB: Characterization of an animal model of ventilator-acquired pneumonia. *Chest* 1999; 115:200–9
7. Charles PE, Etienne M, Croisier D, Piroth L, Lequeu C, Pugin J, Portier H, Chavanet P: The impact of mechanical ventilation on the moxifloxacin treatment of experimental pneumonia caused by *Streptococcus pneumoniae*. *Crit Care Med* 2005; 33:1029–35
8. Whitehead TC, Zhang H, Mullen B, Slutsky AS: Effect of mechanical ventilation on cytokine response to intratracheal lipopolysaccharide. *ANESTHESIOLOGY* 2004; 101:52–8
9. Pugin J, Dunn-Siegrist I, Dufour J, Tissières P, Charles PE, Comte R: Cyclic stretch of human lung cells induces an acidification and promotes bacterial growth. *Am J Respir Cell Mol Biol* 2008; 38:362–70
10. Brégeon F, Roch A, Delpierre S, Ghigo E, Autillo-Touati A, Kajikawa O, Martin TR, Pugin J, Portugal H, Auffray JP, Jammes Y: Conventional mechanical ventilation of healthy lungs induced pro-inflammatory cytokine gene transcription. *Respir Physiol Neurobiol* 2002; 132:191–203
11. Wolthuis EK, Vlaar AP, Choi G, Roelofs JJ, Juffermans NP, Schultz MJ: Mechanical ventilation using non-injurious ventilation settings causes lung injury in the absence of pre-existing lung injury in healthy mice. *Crit Care* 2009; 13:R1
12. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. The Acute Respiratory Distress Syndrome Network. *N Engl J Med* 2000; 342:1301–8
13. Villar J: Low vs high positive end-expiratory pressure in the ventilatory management of acute lung injury. *Minerva Anesthesiol* 2006; 72:357–62
14. Otto CM, Markstaller K, Kajikawa O, Karmrodt J, Syring RS, Pfeiffer B, Good VP, Frevert CW, Baumgardner JE: Spatial and temporal heterogeneity of ventilator-associated lung injury after surfactant depletion. *J Appl Physiol* 2008; 104:1485–94
15. Pavone L, Albert S, DiRocco J, Gatto L, Nieman G: Alveolar instability caused by mechanical ventilation initially damages the nondependent normal lung. *Crit Care* 2007; 11:R104
16. Sartorius A, Lu Q, Vieira S, Tonnellier M, Lenaour G, Goldstein I, Rouby JJ: Mechanical ventilation and lung infection in the genesis of air-space enlargement. *Crit Care* 2007; 11:R14
17. Nahum A, Hoyt J, Schmitz L, Moody J, Shapiro R, Marini JJ: Effect of mechanical ventilation strategy on dissemination of intratracheally instilled *Escherichia coli* in dogs. *Crit Care Med* 1997; 25:1733–43
18. Cakar N, Akinci O, Tugrul S, Ozcan PE, Esen F, Eraksoy H, Cagatay A, Telci L, Nahum A: Recruitment maneuver: Does it promote bacterial translocation? *Crit Care Med* 2002; 30:2103–6
19. Bouadma L, Schortgen F, Ricard JD, Martet G, Dreyfuss D, Saumon G: Ventilation strategy affects cytokine release after mesenteric ischemia-reperfusion in rats. *Crit Care Med* 2004; 32:1563–9
20. Schortgen F, Bouadma L, Joly-Guillou ML, Ricard JD, Dreyfuss D, Saumon G: Infectious and inflammatory dissemination are affected by ventilation strategy in rats with unilateral pneumonia. *Intensive Care Med* 2004; 30:693–701
21. Savel RH, Yao EC, Gropper MA: Protective effects of low tidal volume ventilation in a rabbit model of *Pseudomonas aeruginosa*-induced acute lung injury. *Crit Care Med* 2001; 29:392–8
22. Terragni PP, Rosboch G, Tealdi A, Corno E, Menaldo E, Davini O, Gandini G, Herrmann P, Mascia L, Quintel M, Slutsky AS, Gattinoni L, Ranieri VM: Tidal hyperinflation during low tidal volume ventilation in acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2007; 175:160–6
23. Constantin JM, Grasso S, Chanques G, Afort J, Futier E, Sebbane M, Jung B, Gallix B, Bazin JE, Rouby JJ, Jaber S: Lung morphology predicts response to recruitment maneuver in patients with acute respiratory distress syndrome. *Crit Care Med* 2010; 38:1108–17
24. Charles PE, Martin L, Etienne M, Croisier D, Piroth L, Lequeu C, Pugin J, Portier H, Chavanet P: Influence of positive end-expiratory pressure (PEEP) on histopathological and bacteriological aspects of pneumonia during low tidal volume mechanical ventilation. *Intensive Care Med* 2004; 30:2263–70
25. Kurahashi K, Ota S, Nakamura K, Nagashima Y, Yazawa T, Satoh M, Fujita A, Kamiya R, Fujita E, Baba Y, Uchida K, Morimura N, Andoh T, Yamada Y: Effect of lung-protective ventilation on severe *Pseudomonas aeruginosa* pneumonia and sepsis in rats. *Am J Physiol Lung Cell Mol Physiol* 2004; 287:L402–10
26. Blanch L, Mancebo J, Perez M, Martinez M, Mas A, Betbese AJ, Joseph D, Ballús J, Lucangelo U, Bak E: Short-term effects of prone position in critically ill patients with acute respiratory distress syndrome. *Intensive Care Med* 1997; 23:1033–9
27. Galiatsou E, Kostanti E, Svarna E, Kitsakos A, Koulouras V, Efremidis SC, Nakos G: Prone position augments recruitment and prevents alveolar overinflation in acute lung injury. *Am J Respir Crit Care Med* 2006; 174:187–97
28. Broccard A, Shapiro RS, Schmitz LL, Adams AB, Nahum A, Marini JJ: Prone positioning attenuates and redistributes ventilator-induced lung injury in dogs. *Crit Care Med* 2000; 28:295–303
29. Valenza F, Guglielmi M, Maffioletti M, Tedesco C, Maccagni P, Fossali T, Aletti G, Porro GA, Irace M, Carlesso E, Carboni N, Lazzerini M, Gattinoni L: Prone position delays the progression of ventilator-induced lung injury in rats: Does lung strain distribution play a role? *Crit Care Med* 2005; 33:361–7
30. Nakos G, Batistatou A, Galiatsou E, Konstanti E, Koulouras V, Kanavaros P, Doulis A, Kitsakos A, Karachaliou A, Lekka ME, Bai M: Lung and ‘end organ’ injury due to mechanical ventilation in animals: Comparison between the prone and supine positions. *Crit Care* 2006; 10:R38
31. Richard JC, Bregeon F, Costes N, Bars DL, Tourvieille C, Lavenne F, Janier M, Bourdin G, Gimenez G, Guerin C: Effects of prone position and positive end-expiratory pressure on lung perfusion and ventilation. *Crit Care Med* 2008; 36:2373–80
32. Goldstein I, Bughalo MT, Marquette CH, Lenaour G, Lu Q, Rouby JJ; Experimental ICU Study Group: Mechanical ventilation-induced air-space enlargement during experimental pneumonia in piglets. *Am J Respir Crit Care Med* 2001; 163:958–64
33. Brackenburg AM, McCaig LA, Yao LJ, Veldhuizen RA, Lewis JF: Host response to intratracheally instilled bacteria in ventilated and nonventilated rats. *Crit Care Med* 2004; 32:2502–7
34. Croisier D, Etienne M, Bergoin E, Charles PE, Lequeu C, Piroth L, Portier H, Chavanet P: Mutant selection window in levofloxacin and moxifloxacin treatments of experimental pneumococcal pneumonia in a rabbit model of human therapy. *Antimicrob Agents Chemother* 2004; 48:1699–707
35. Gattinoni L, Pelosi P, Vitale G, Pesenti A, D’Andrea L, Mascheroni D: Body position changes redistribute lung computed-tomographic density in patients with acute respiratory failure. *ANESTHESIOLOGY* 1991; 74:15–23
36. Pelosi P, Bottino N, Chiumello D, Caironi P, Panigada M, Gamberoni C, Colombo G, Bigatello LM, Gattinoni L: Sigh in supine and prone position during acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2003; 167:521–7
37. Guerin C, Gaillard S, Lemasson S, Ayzac L, Girard R, Beuret P, Palmier B, Le QV, Sirodot M, Rosselli S, Cadiergue V, Sainy JM, Barbe P, Combout E, Debatty D, Rouffineau J, Ezingeard E, Millet O, Guelon D, Rodriguez L, Martin O, Renault A, Sibille JP, Kaidomar M: Effects of systematic prone positioning in hypoxemic acute respiratory failure: A randomized controlled trial. *JAMA* 2004; 292:2379–87
38. Beuret P, Carton MJ, Nouridine K, Kaaki M, Tramon G, Ducreux JC: Prone position as prevention of lung injury in

- comatose patients: A prospective, randomized, controlled study. *Intensive Care Med* 2002; 28:564-9
39. Sud S, Friedrich JO, Taccone P, Polli F, Adhikari NK, Latini R, Pesenti A, Guérin C, Mancebo J, Curley MA, Fernandez R, Chan MC, Beuret P, Voggenreiter G, Sud M, Tognoni G, Gattinoni L: Prone ventilation reduces mortality in patients with acute respiratory failure and severe hypoxemia: Systematic review and meta-analysis. *Intensive Care Med* 2010; 36:585-99
 40. Villar J, Cabrera N, Casula M, Flores C, Valladares F, Muros M, Blanch L, Slutsky AS, Kacmarek RM: Mechanical ventilation modulates Toll-like receptor signaling pathway in a sepsis-induced lung injury model. *Intensive Care Med* 2010; 36:1049-57
 41. Charles PE, Barbar SD: Toll-like receptors: A link between mechanical ventilation, innate immunity and lung injury? *Intensive Care Med* 2010; 36:909-11
 42. Pelosi P, Tubiolo D, Mascheroni D, Vicardi P, Crotti S, Valenza F, Gattinoni L: Effects of the prone position on respiratory mechanics and gas exchange during acute lung injury. *Am J Respir Crit Care Med* 1998; 157:387-93
 43. Oudin S, Pugin J: Role of MAP kinase activation in interleukin-8 production by human BEAS-2B bronchial epithelial cells submitted to cyclic stretch. *Am J Respir Cell Mol Biol* 2002; 27:107-14
 44. Panigada M, Berra L, Greco G, Stylianou M, Kolobow T: Bacterial colonization of the respiratory tract following tracheal intubation-effect of gravity: An experimental study. *Crit Care Med* 2003; 31:729-37
 45. Joliet P, Bulpa P, Chevrolet JC: Effects of the prone position on gas exchange and hemodynamics in severe acute respiratory distress syndrome. *Crit Care Med* 1998; 26:1977-85

ANESTHESIOLOGY REFLECTIONS FROM THE WOOD LIBRARY-MUSEUM

Statikil Sparked Paul Wood's Interest



Fifty years in the wake of losing Dr. Paul Meyer Wood, we celebrate the semicentennial of his namesake library-museum in Park Ridge, Illinois. Among the many odd items Dr. Wood collected was this bottle of Statikil, which was distributed by a firm located just 1.5 miles southwest of the New York brownstone that once harbored the Wood Library-Museum. Product labeling claimed that any garment immersed in Statikil (and then squeezed out and hung out to dry) would be rendered "spark-proof." Statikil was yet another weapon against static electrical ignition of flammable anesthetic gases in the operating room. (Copyright © the American Society of Anesthesiologists, Inc.)

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